NIH-3T3/Human IGF-1 R Stable Cell Line Development Service

Catalog No.	Size
CNIH-ATP102	2 × (1 vial contains ~5×10^6 cells)

• *Description*

NIH-3T3/Human IGF-1 R Stable Cell Line was engineered to express the receptor full length human IGF-1 R (Gene ID: 3480). Surface expression of Human IGF-1 R was confirmed by flow cytometry.

• *Application*

- Useful for cell-based IGF-1 R binding assay

• *Cell Line Profile*

Cell line	NIH-3T3/Human IGF-1 R Stable Cell Line
Host Cell	NIH-3T3
Property	Adherent
Complete Growth Medium	DMEM + 10% NBCS
Selection Marker	Hygromycin (20 μg/mL)
Incubation	37°C with 5% CO_2
Doubling Time	22-24 hours
Transduction Technique	Lentivirus

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• *Materials Required for Cell Culture*

- DMEM medium (Gibco, Cat.No.11965-092)
- Newborn calf serum (Gibco, Cat.No.16010-159)
- Hygromycin B (Invitrogen, Cat.No.10687010)
- Complete Growth Medium: DMEM + 10% NBCS
- Culture Medium: DMEM + 10% NBCS, Hygromycin (20 μg/mL)
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- MONWEI^{TM}SmartCell200 (Shanghai Monwei Biomedical Technology Co. ,Ltd)
- CO_2 Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)

• *Recovery*

1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
4. Discard the supernatant and resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
5. Incubate at 37°C with 5% CO_2 incubator.

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• *Subculture*

1. Remove and discard culture medium.
2. Wash the cells once with sterile PBS.
3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-5 minutes, until 90% of the cells have detached
4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
5. Add appropriate aliquots of the cell suspension to new culture vessel.
6. Incubate at 37°C with 5% CO_2 incubator.

- **Subcultivation Ratio:** A subcultivation ratio of 1:4 to 1:8 is recommended.
- **Medium Renewal:** Every 2 to 3 days.

• *Cryopreservation*

1. Remove and discard spent medium.
2. Detach cells from the cell culture flasks with 0.25% trypsin.
3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
4. Resuspend the cell pellets with complete growth medium and count viable cells.
5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of 5×10^6 to 1×10^7 cells/mL.
6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in a –80°C freezer overnight, then transferring to liquid nitrogen storage.

• *Storage*

- **Product format:** Frozen
- **Storage conditions:** Liquid nitrogen immediately upon receipt

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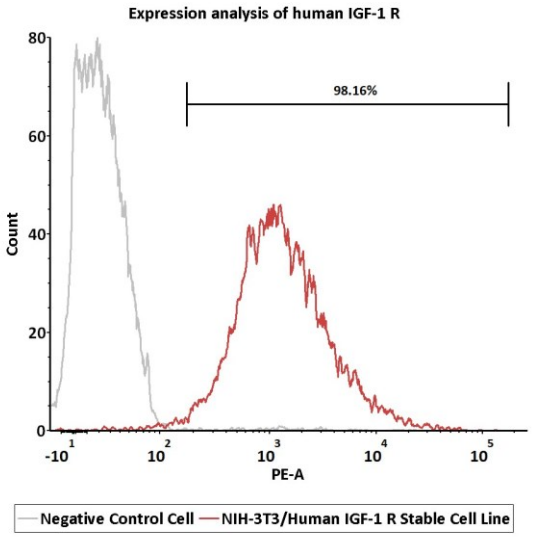
• *Receptor Assay*



Catalog No.	Stable Cell Line	MFI for IGF-1 R (PE)
NA	Negative Control Cell	17.93
CNIH-ATP102	NIH-3T3/Human IGF-1 R Stable Cell Line Development Service	1178.26

Fig1. Expression analysis of human IGF-1 R on NIH-3T3/Human IGF-1 R Stable Cell Line by FACS. Cell surface staining was performed on NIH-3T3/Human IGF-1 R Stable Cell Line or negative control cell using PE-labeled anti-human IGF-1 R antibody.

• *License Disclosure*

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