

# Human TGF-beta R (Luc) HEK293 Reporter Cell

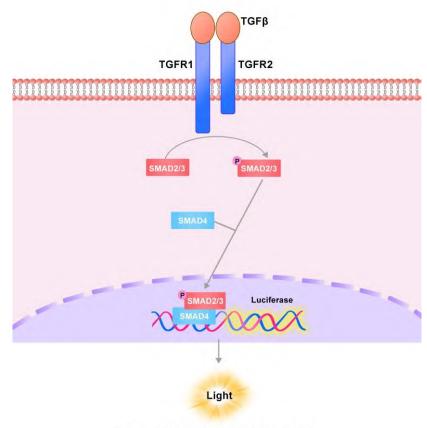
Catalog No.	Size
CHEK-ATF145	$2 \times (1 \text{ vial contains } \sim 5 \times 10^{6} \text{ cells})$

#### • Description

The Human TGF-beta R (Luc) HEK293 Reporter Cell was engineered to express Smad signaling response element driving luciferase expressing systems. When stimulated with TGF-beta (TGF-beta 1/2/3) protein, receptor-mediated signaling can drive Smad-mediated luminescence. Neutralization of biological effect of the ligand-receptor interaction by corresponding antibody results in a decrease in luminescence.

#### • Application

• Screen for neutralizing antibodies blocking the ligand-receptor interaction.



Human TGF-beta R (Luc) HEK293 Reporter Cell



### • Cell Line Profile

Cell line	
Host Cell	
Property	
Complete Growth Medium	
Selection Marker	
Incubation	
Doubling Time	
Transduction Technique	

Human TGF-beta R (Luc) HEK293 Reporter Cell HEK293 Adherent DMEM + 10% FBS Puromycin (2 µg/mL) 37°C with 5% CO<sub>2</sub> 22-24 hours Lentivirus

### • Materials Required for Cell Culture

- DMEM medium (Gibco, Cat.No.11965-092)
- Fetal bovine serum (CellMax, Cat.No.SA211.02)
- Puromycin (InvivoGen, Cat.No.ant-pr-5b)
- 0.25% Trypsin-EDTA (1X), Phenol Red (Gibco, Cat.No.25200-056)
- Penicillin-Streptomycin (Gibco, Cat.No.15140-122)
- Phosphate Buffered Saline (1X) (HyClone, Cat.No.SH30256.01)
- Complete Growth Medium: DMEM + 10% FBS
- Culture Medium: DMEM + 10% FBS, Puromycin (2 µg/mL)
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- Luna cell counter (Logos Biosystems, LUNA-II)
- CO<sub>2</sub> Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)



#### • Recovery

- 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
- 3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
- 4. Resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
- 5. Incubate at 37°C with 5% CO<sub>2</sub> incubator until the cells are ready to be split.

#### • Subculture

- 1. Remove and discard culture medium.
- 2. Wash the cells once with sterile PBS.
- 3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
- 4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessel.
- 6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

Subcultivation Ratio: A subcultivation ratio of 1:6 to 1:10 is recommended.

Medium Renewal: Every 2 to 3 days.

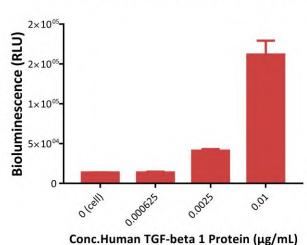


### • Cryopreservation

- 1. Remove and discard spent medium.
- 2. Detach cells from the cell culture flasks with 0.25% trypsin.
- 3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
- 4. Resuspend the cell pellets with complete growth medium and count viable cells.
- 5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of  $5 \times 10^6$  to  $1 \times 10^7$  cells/mL.
- 6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in a -80°C freezer overnight, then transferring to liquid nitrogen storage.
- Storage
  - **Product format:** Frozen
  - Storage conditions: Liquid nitrogen immediately upon receipt



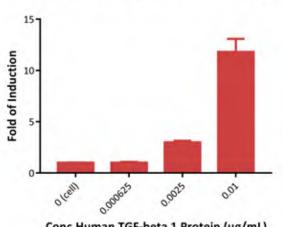
### • Signaling Bioassay



Human TGF-beta 1 Protein Stimulation (RLU)

Conc.Human (GF-beta 1 Protein (µg/mL)

**Fig1. Response to human TGF-beta 1 protein (RLU).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 1 protein (Cat.No.TG1-H4212). The max induction fold was approximately 11.81.

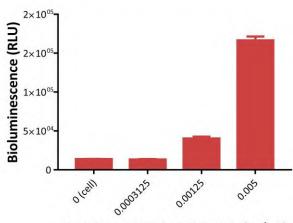


Human TGF-beta 1 Protein Stimulation (FOLD)

Conc.Human TGF-beta 1 Protein (µg/mL)

**Fig2. Response to human TGF-beta 1 protein (FOLD).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 1 protein (Cat.No.TG1-H4212). The max induction fold was approximately 11.81.

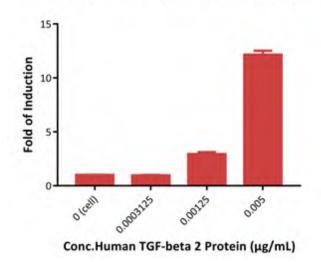




#### Human TGF-beta 2 Protein Stimulation (RLU)

Conc.Human TGF-beta 2 Protein (µg/mL)

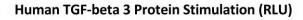
**Fig3. Response to human TGF-beta 2 protein (RLU).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 2 protein (Cat.No.TG2-H4215). The max induction fold was approximately 12.15.

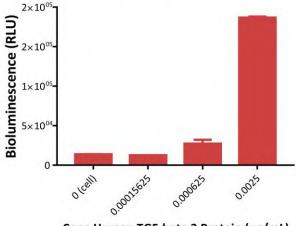


Human TGF-beta 2 Protein Stimulation (FOLD)

**Fig4. Response to human TGF-beta 2 protein (FOLD).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 2 protein (Cat.No.TG2-H4215). The max induction fold was approximately 12.15.

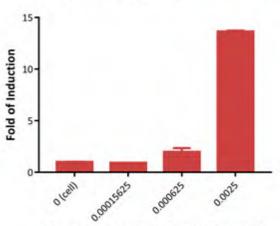






Conc.Human TGF-beta 3 Protein (µg/mL)

**Fig5. Response to human TGF-beta 3 protein (RLU).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 3 protein (Cat.No.TG3-H5213). The max induction fold was approximately 13.64.



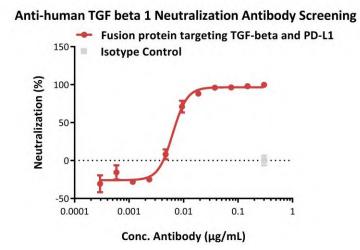
### Human TGF-beta 3 Protein Stimulation (FOLD)

Conc.Human TGF-beta 3 Protein (µg/mL)

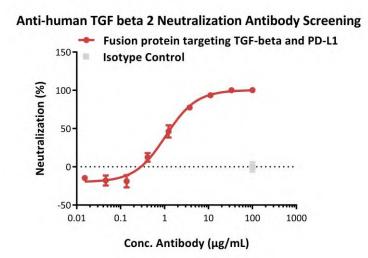
**Fig6. Response to human TGF-beta 3 protein (FOLD).** The Human TGF-beta R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human TGF-beta 3 protein (Cat.No.TG3-H5213). The max induction fold was approximately 13.64.



### Application

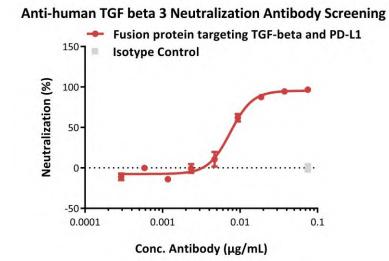


**Fig7.** Inhibition of human TGF-beta 1 protein protein-induced reporter activity by a bifunctional fusion protein targeting TGF-beta and PD-L1. This reporter cell was incubated with serial dilutions of the bifunctional fusion protein in the presence of human TGF-beta 1 protein (Cat.No.TG1-H4212) with a final concentration of 0.005 μg/mL. The EC50 of the bifunctional fusion protein (Bintrafusp alfa) is approximately 0.006323 μg/mL.



**Fig8.** Inhibition of human TGF-beta 2 protein protein-induced reporter activity by a bifunctional fusion protein targeting TGF-beta and PD-L1. This reporter cell was incubated with serial dilutions of the bifunctional fusion protein in the presence of human TGF-beta 2 protein (Cat.No.TG2-H4215) with a final concentration of 0.003 μg/mL. The EC50 of the bifunctional fusion protein (Bintrafusp alfa) is approximately 1.06 μg/mL.





**Fig9.** Inhibition of human TGF-beta 3 protein protein-induced reporter activity by a bifunctional fusion protein targeting TGF-beta and PD-L1. This reporter cell was incubated with serial dilutions of the bifunctional fusion protein in the presence of human TGF-beta 3 protein (Cat.No.TG3-H5213) with a final concentration of 0.002 µg/mL. The EC50 of the bifunctional fusion protein (Bintrafusp alfa) is approximately 0.0075 µg/mL.

#### • License Disclosure

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#### • Related Products

Products	<u>Cat.No.</u>
Human TGF-Beta 1/TGFB1 Protein, premium grade	TG1-H4212
Human TGF-Beta 2/TGFB2 Protein, Tag Free	TG2-H4215
Human TGF-beta 3/TGFB3 Protein, premium grade	TG3-H5213