

### TCF/LEF (Luc) HEK293 Reporter Cell

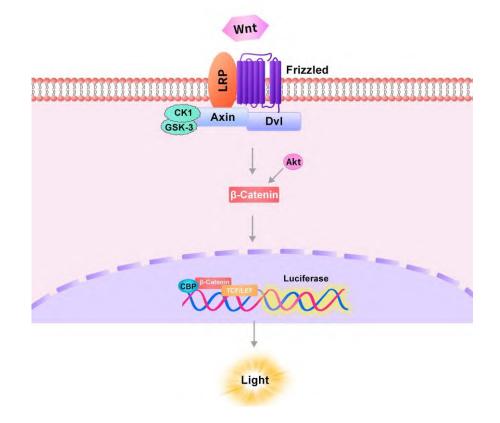
Catalog No.	Size
CHEK-ATF114	$2 \times (1 \text{ vial contains } \sim 5 \times 10^{6} \text{ cells})$

#### • Description

The TCF/LEF (Luc) HEK293 Reporter Cell was engineered to express TCF/LEF signaling response element driving luciferase expressing systems, designed for monitoring the activity of the Wnt/β-catenin signaling pathway. When stimulated with human Wnt protein, receptor-mediated signaling can drive TCF/LEF-mediated luminescence. Inhibition of biological effect of human Wnt protein by corresponding inhibitors results in a decrease in luminescence.

#### • Application

- Monitor Wnt signaling pathway activity.
- Screen for Wnt/β-catenin targeted agents.





### • Cell Line Profile

Cell line	
Host Cell	
Property	
Complete Growth Medium	
Selection Marker	
Incubation	
Doubling Time	
Transduction Technique	

TCF/LEF (Luc) HEK293 Reporter Cell HEK293 Adherent DMEM + 10% FBS Puromycin (2 µg/mL) 37°C with 5% CO<sub>2</sub> 22-24 hours Lentivirus

### • Materials Required for Cell Culture

- DMEM medium (Gibco, Cat.No.11965-092)
- Fetal bovine serum (CellMax, Cat.No.SA211.02)
- Puromycin (InvivoGen, Cat.No.ant-pr-5b)
- Complete Growth Medium: DMEM + 10% FBS
- Culture Medium: DMEM + 10% FBS, Puromycin (2 µg/mL)
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- Luna cell counter (Logos Biosystems, LUNA-II)
- CO<sub>2</sub> Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)



#### • Recovery

- 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
- 3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
- 4. Resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
- 5. Incubate at 37°C with 5% CO<sub>2</sub> incubator until the cells are ready to be split.

#### • Subculture

- 1. Remove and discard culture medium.
- 2. Wash the cells once with sterile PBS.
- 3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
- 4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessel.
- 6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

Subcultivation Ratio: A subcultivation ratio of 1:6 to 1:10 is recommended.

Medium Renewal: Every 2 to 3 days.

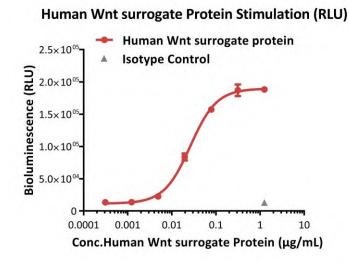


### • Cryopreservation

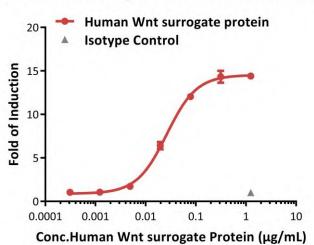
- 1. Remove and discard spent medium.
- 2. Detach cells from the cell culture flasks with 0.25% trypsin.
- 3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
- 4. Resuspend the cell pellets with complete growth medium and count viable cells.
- 5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of  $5 \times 10^6$  to  $1 \times 10^7$  cells/mL.
- 6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in  $a 80^{\circ}$ C freezer overnight, then transferring to liquid nitrogen storage.
- Storage
  - **Product format:** Frozen
  - Storage conditions: Liquid nitrogen immediately upon receipt



#### • Signaling Bioassay



**Fig1.Response to human Wnt surrogate protein (RLU).** This reporter cell was incubated with serial dilutions of human Wnt surrogate protein. The EC<sub>50</sub> was approximately 0.02628  $\mu$ g/mL.



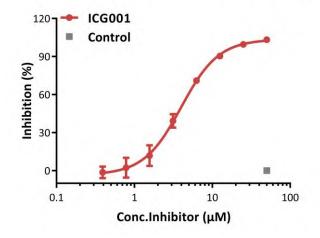
#### Human Wnt surrogate Protein Stimulation (FOLD)

**Fig2.Response to human Wnt surrogate protein (FOLD).** This reporter cell was incubated with serial dilutions of human Wnt surrogate protein. The max induction fold was approximately 14.



### • Application

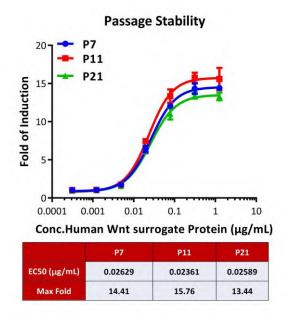
### Wnt/β-catenin pathway Inhibitor Screening



**Fig3.** Inhibition of human Wnt surrogate protein-induced reporter activity by Wnt/β-catenin pathway Inhibitor. This reporter cell was incubated with serial dilutions of inhibitors in the presence of human Wnt surrogate protein with a final concentration of 0.050 µg/mL. The EC<sub>50</sub> of Wnt/β-catenin pathway Inhibitor (ICG001) was approximately 3.993 µM.



#### • Passage Stability



**Fig4.** Passage stability analysis by Signaling Bioassay. The continuously growing TCF/LEF (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of human Wnt surrogate protein. Human Wnt surrogate protein stimulated response demonstrates passage stabilization (fold induction and EC50) across passage 7-21.

#### • License Disclosure

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