

Human GLP-1R (Luc) HEK293 Reporter Cell

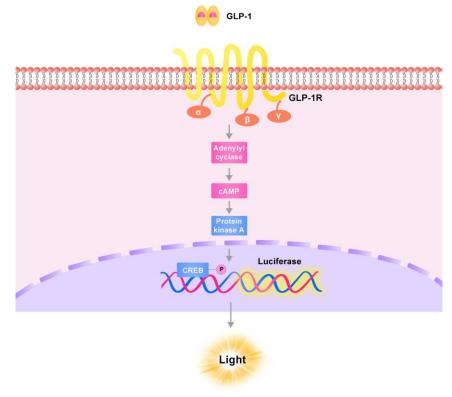
Catalog No.	Size
CHEK-ATF096	$2 \times (1 \text{ vial contains } \sim 5 \times 10^{6} \text{ cells})$

• Description

The Human GLP-1R (Luc) HEK293 Reporter Cell was engineered to not only express CREB signaling response element, but also express the receptor full length human GLP-1R (Gene ID: 2740), which can drive luciferase expressing systems by GLP-1R agonists or glucagon-like peptide 1 (GLP-1) stimulation. In the absence of agonist or GLP-1, the GLP-1R receptor is not activated and luminescence signal is low. In the presence of agonist or GLP-1, the GLP-1R pathway-activated luminescence can be detected in a dose-dependent manner.

• Application

Screen for agonists that can bind and activate GLP-1R.



Human GLP-1R (Luc) HEK293 Reporter Cell



• Cell Line Profile

Cell line	Human GLP-1R (Luc) I
Host Cell	HEK
Property	Adhe
Complete Growth Medium	DMEM +
Selection Marker	Puromycin (2 µg/mL) + H
Incubation	37°C with
Doubling Time	22-24
Transduction Technique	Lentiv

Human GLP-1R (Luc) HEK293 Reporter Cell HEK293 Adherent DMEM + 10% FBS Puromycin (2 µg/mL) + Hygromycin (20 µg/mL) 37°C with 5% CO₂ 22-24 hours Lentivirus

• Materials Required for Cell Culture

- DMEM medium (Gibco, Cat.No.11965-092)
- Fetal bovine serum (CellMax, Cat.No.SA211.02)
- Puromycin (InvivoGen, Cat.No.ant-pr-5b)
- Hygromycin B (Invitrogen, Cat.No.10687010)
- Complete Growth Medium: DMEM + 10% FBS
- Culture Medium: DMEM + 10% FBS, Hygromycin (20 µg/mL), Puromycin (2 µg/mL)
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- Luna cell counter (Logos Biosystems, LUNA- II)
- CO₂ Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)



• Recovery

- 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
- 3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
- 4. Resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
- 5. Incubate at 37° C with 5% CO₂ incubator until the cells are ready to be split.

• Subculture

- 1. Remove and discard culture medium.
- 2. Wash the cells once with sterile PBS.
- 3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
- 4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessel.
- 6. Incubate at 37 $^\circ\!\mathrm{C}$ with 5% CO_2 incubator.

Subcultivation Ratio: A subcultivation ratio of 1:6 to 1:10 is recommended.

Medium Renewal: Every 2 to 3 days.



• Cryopreservation

- 1. Remove and discard spent medium.
- 2. Detach cells from the cell culture flasks with 0.25% trypsin.
- 3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
- 4. Resuspend the cell pellets with culture medium and count viable cells.
- 5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of 5×10^6 to 1×10^7 cells/mL.
- 6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in $a 80^{\circ}$ C freezer overnight, then transferring to liquid nitrogen storage.
- Storage
 - **Product format:** Frozen
 - Storage conditions: Liquid nitrogen immediately upon receipt



• Receptor Assay

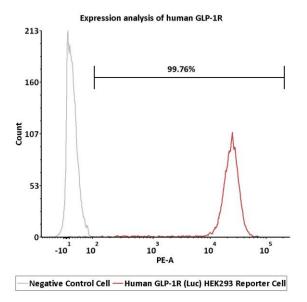


Fig1. Expression analysis of human GLP-1R on Human GLP-1R (Luc) HEK293 Reporter Cell by FACS. Cell surface staining was performed on Human GLP-1R (Luc) HEK293 Reporter Cell or negative control cell using PE-labeled anti-human GLP-1R antibody.

• Application

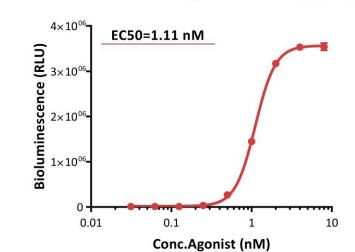


Fig2. Bioactivity analysis of human GLP-1R agonist (RLU). This reporter cell was incubated with serial dilutions of Tirzepatide (a dual GLP-1R and GIPR agonist). The EC50 of Tirzepatide was approximately 1.11 nM.

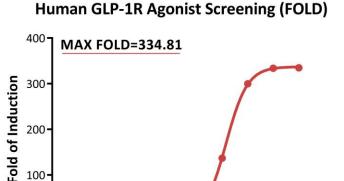
Human GLP-1R Agonist Screening (RLU)

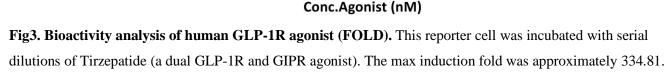


200

100

0 0.01



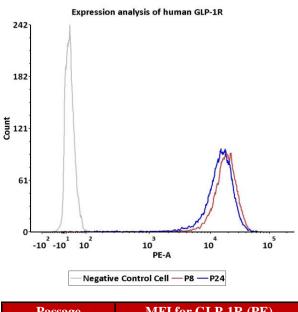


1

10

0.1

Passage Stability



Passage	MFI for GLP-1R (PE)
P8	17421
P24	14536

Fig4. Passage stability analysis of receptor expression by FACS. Flow cytometry surface staining of human GLP-1R on Human GLP-1R (Luc) HEK293 Reporter Cell demonstrates consistent mean fluorescent intensity across passage 8-24.



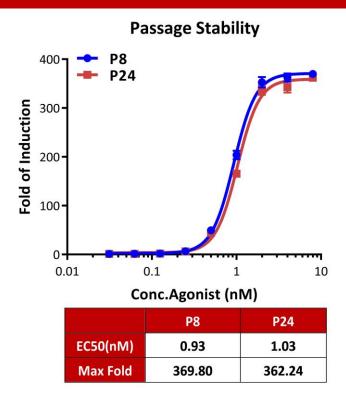


Fig5. Passage stability analysis by Signaling Bioassay. The continuously growing Human GLP-1R (Luc) HEK293 Reporter Cell was stimulated with serial dilutions of Tirzepatide (a dual GLP-1R and GIPR agonist). Tirzepatide stimulated response demonstrates passage stabilization (fold induction and EC50) across passage 8-24.

• License Disclosure

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• Related Products

Products

Human GCGR (Luc) HEK293 Reporter Cell Human GIPR (Luc) HEK293 Reporter Cell <u>Cat.No.</u> CHEK-ATF103 CHEK-ATF104