ClinMax™ Human IFN-γ ELISA Kit, PRO

Catalog Number：CEA-C006

Assay Tests: 96 tests

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures

IMPORTANT: Please carefully read this user guide before performing your experiment.

Product information

This kit is specifically designed for the accurate quantitation of human Interferon Gamma (IFN-γ) from cell culture supernates, serum and plasma.

The principle of this assay employs a quantitative sandwich enzyme immunoassay approach. Initially, a microplate is coated with a capture antibody. Then, samples and biotinylated capture antibody are added to the wells. After the removal of any unbound materials through washing, streptavidin-HRP (SA-HRP) conjugate is added to the wells. Streptavidin has a very high affinity for biotin, so it binds to the biotinylated capture antibody that is already bound to the target antigen. After washing, a substrate specific to HRP is added to the wells. HRP catalyzes a reaction that converts the substrate into a detectable signal, often a color change or luminescence, depending on the substrate used. This enzymatic reaction amplifies the signal, allowing for higher sensitivity in detecting the target analyte. The intensity of the signal is measured using a spectrophotometer.

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NOTE:

1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.
2. Please do not use the kit after the expiration date indicated on the kit label.
3. Do not mix or substitute reagents with those from other lots or sources.

Manufactured and distributed by
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Distributed by:
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Contents

The kit contains sufficient reagents for 96 wells.

Catalog	Contents	Amount
CEA006-C01	Pre-coated Anti-IFN-γ Ab Microplate	1 plate
CEA006-C02	IFN-γ Calibrator	14 μg ×2
CEA006-C03	Biotin-Anti-IFN-γ Ab Con. Solution	150 μL
CEA006-C04	Biotin-Ab Dilution Buffer	8 mL
CEA006-C05	IFN-γ SA-HRP Con. Solution	0.5 mL
CEA006-C06	SA-HRP Dilution Buffer	15 mL
CEA006-C07	20× Washing Buffer	50 mL
CEA006-C08	Sample Dilution Buffer	15 mL ×2
CEA006-C09	Substrate Solution	12 mL
CEA006-C10	Stop Solution	6 mL

Storage

Keep the unopened kit stored at 2-8 °C. Avoid using the kit beyond its expiration date. For opened kit and reconstituted reagents, with the exception of the two contents listed in following table, others can be stored for up to 30 days at 2-8 °C.

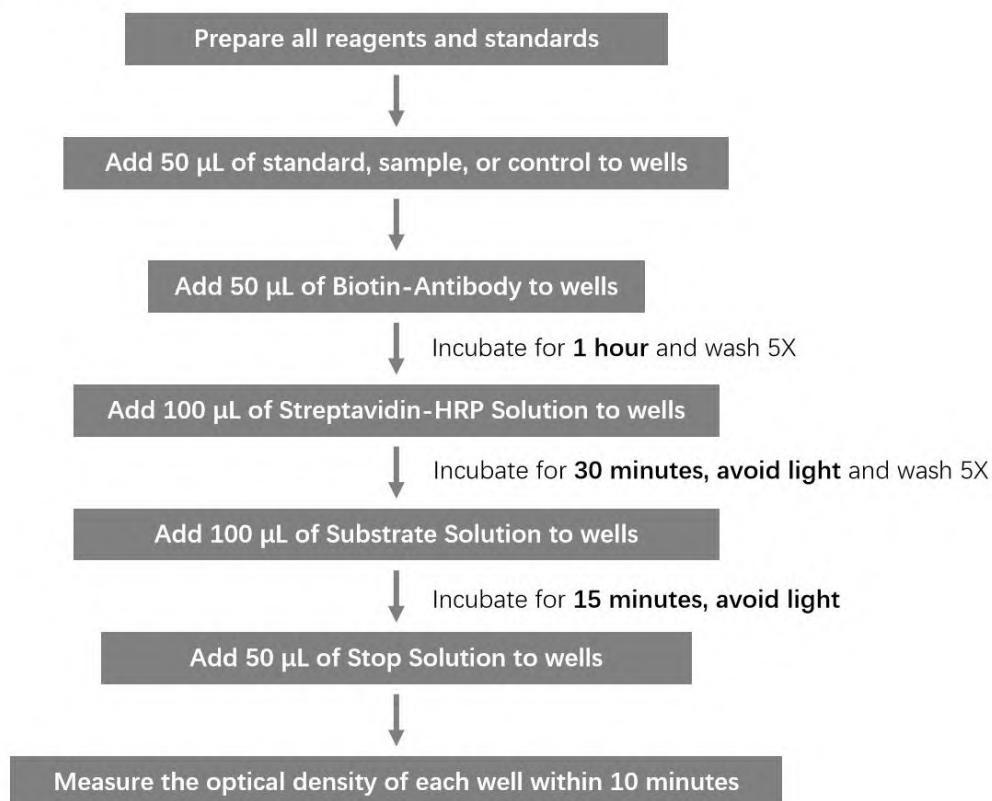| Contents	Storage conditions
Pre-coated Anti-IFN-γ Ab Microplate	Return unused wells to the foil pouch, reseal along entire edge. May be stored for up to 1 month at 2-8°C.
IFN-γ Calibrator	Aliquot and store for up to 1 month at -70°C in a freezer. Avoid repeated freeze-thaw cycles.

NOTE: Streptavidin-HRP Con. Solution and Substrate Solution should avoid light.

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Required materials not supplied.

Instrument	Microplate reader capable of measuring absorbance at 450 nm
Reagents	Deionized, ultrapure or distilled water
Consumables	50 mL and 500 mL graduated cylinders
	Pipettes and pipette tips
	Tubes to prepare standard dilutions.

Workflow

Analyte: IFN-γ

Prepare all reagents and standards

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Add 50 µL of standard, sample, or control to wells

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Add 50 µL of Biotin-Antibody to wells

↓ Incubate for **1 hour** and wash 5X

Add 100 µL of Streptavidin-HRP Solution to wells

↓ Incubate for **30 minutes, avoid light** and wash 5X

Add 100 µL of Substrate Solution to wells

↓ Incubate for **15 minutes, avoid light**

Add 50 µL of Stop Solution to wells

↓

Measure the optical density of each well within 10 minutes

NOTE：Incubation temperature is 18 ℃-25 ℃

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Prepare the working buffers and standard dilutions.

IMPORTANT: Bring all reagents to room temperature before use. If crystals have formed in buffer solution, place the buffer solution in an 37°C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

Prepare the working buffers.

1.   1×Washing Buffer: Dilute 50 mL 20×Washing Buffer with deionized or distilled water to 1000 mL.

2.   Biotin-Anti-IFN-γ Ab Solution: Add 120 μL of Biotin-Anti-IFN-γ Ab Con. Solution to 6 mL Biotin-Ab Dilution Buffer, thoroughly mix. The solution was freshly prepared just before use.

3.   IFN-γ SA-HRP Solution: Add 400 μL of IFN-γ SA-HRP Con. Solution to 12 mL of SA-HRP Dilution Buffer, thoroughly mix. The solution was freshly prepared just before use.

Prepare the reconstituted Calibrator.

Add 0.1 mL ultrapure water to the provided lyophilized product（CEA006-C02），dissolve at room temperature for 15-30 minutes, and mix by gently pipetting. The concentration of reconstituted human IFN-γ Calibrator is 140 μg /mL.

NOTE: Avoiding vigorous shaking or vortexing. The reconstituted solution should be stored at -70°C. The freeze-thaw cycle should not exceed 1 time, and the size of the aliquot should not be less than 5 μg.

Prepare the standard serial dilutions.

1.   Label a tube "**Cm**". Add 5 μL of the reconstituted human IFN-γ Calibrator and 1995 μL of Sample Dilution Buffer to tube Cm, gently mix well.

2.   Label 8 tubes, one for each standard point: Cal.-1, Cal.-2, Cal.-3, Cal.-4, Cal.-5, Cal.-6, Cal.-7, Cal.-8.

3.   Add 10 μL of the liquid from **Cm** and 3490 μL of Sample Dilution Buffer to tube Cal.-1, thoroughly mix (Cal.-1 =1000 pg/mL).

4.   Prepare serial dilutions for the calibration curve as follows: Add 500 μL of Sample Dilution Buffer to each tube (Cal.-2, Cal.-3, Cal.-4, Cal.-5, Cal.-6, Cal.-7, Cal.-8).

5.   Transfer 500 μL of liquid from Cal.-1 to the tube Cal.-2, and thoroughly mix (Cal.-2 = 500 pg/mL).

6.   Continue to transfer 500 μL of liquid from previous dilution tube to the next dilution tube until add liquid to tube Cal.-8 (7.81 pg/mL).

7.   Sample Dilution Buffer serves as zero standard (blank).
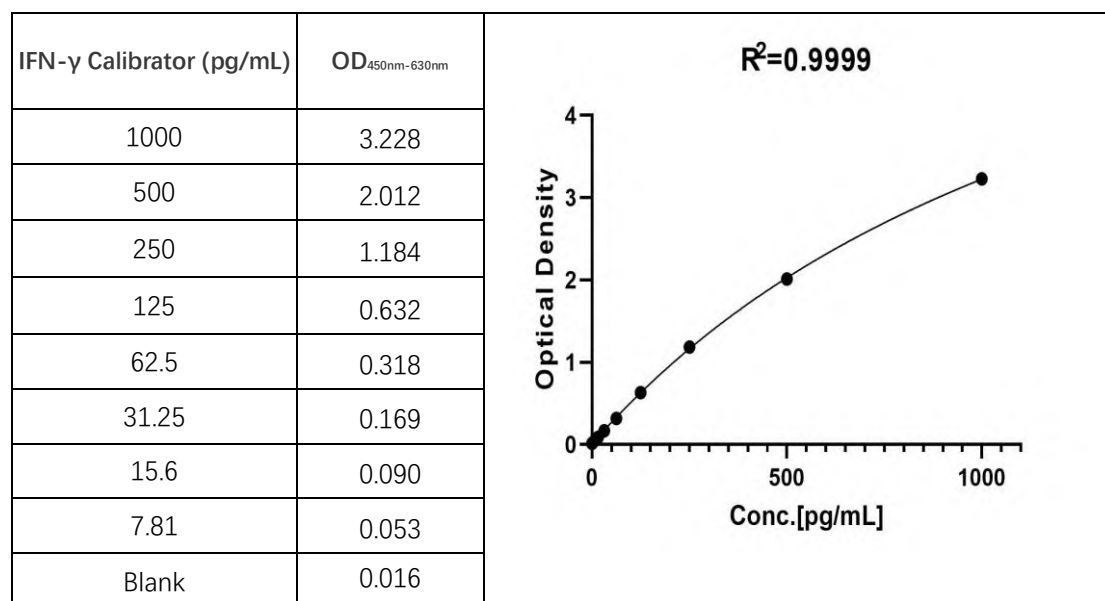
PROCEDURE OF ASSAY

1. Add 50 µL of IFN-γ Calibrator, sample, or control to wells.

2. Add 50 µL Biotin-Anti-IFN-γ Ab Solution to each well, Seal the plate with microplate sealing film. Incubate at room temperature (18-25 °C) for **1 hours.**

3. Aspirate each well and add 300 µL of 1×Washing Buffer to each well, gently tap the plate for **1 minute**. Remove any remaining Washing Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels. Repeat the wash process four times for a total of five washes.

4. Add 100 µL of IFN-γ SA-HRP Solution to each well. Seal the plate with microplate sealing film. Incubate at room temperature (18-25 °C) for **30 minutes, avoid light.**

5. Repeat step 3.

6. Add 100 µL of Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at room temperature (18-25 °C) for **15 minutes, avoid light**.

7. Add 50 µL of Stop Solution to each well. Tap the plate gently to ensure thorough mixing.
   Note: the color in the wells should change from blue to yellow.

8. Read the absorbance at 450nm and 630nm using Microplate reader within 10minutes.
   Note: To reduce the background noise, subtract the readings at 630nm from the readings at 450nm.

CALCULATION OF RESULTS

1. Compute the average of the duplicated readings for every standard, control, and sample. Then, subtract the average optical density (O.D.) of the zero standard(blank).
2. Establish a calibration curve by processing the data using computer software capable of executing a four-parameter logistic (4-PL) curve fitting.
3. Normal range of calibration curve: $R^2 \geqslant 0.9900$.
4. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with dilution buffer and assay repeated.

Typical data

Note: *For each experiment, a calibration curve needs to be set for each microplate, and the specific OD value may vary depending on different laboratories, testers, or equipment. The following example data is for reference only. The sample concentration was calculated based on the results of the calibration curve.*

IFN-γ Calibrator (pg/mL)	OD$_{450nm-630nm}$
1000	3.228
500	2.012
250	1.184
125	0.632
62.5	0.318
31.25	0.169
15.6	0.090
7.81	0.053
Blank	0.016

$R^2=0.9999$

PERFORMANCE CHARACTERISTICS

1. Sensitivity

The minimum detectable concentration (MDC) of IFN-γ is typically less than 4.0 pg/mL. The MDC was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

2. Linearity

Three samples (Serum) spiked with high concentrations of 1800 pg/mL, 1600 pg/mL and 1500 pg/mL were serially diluted with dilution buffer to produce samples with values within the dynamic range of the assay and then assayed. The average recovery of IFN-γ for serum samples is 99.16%.
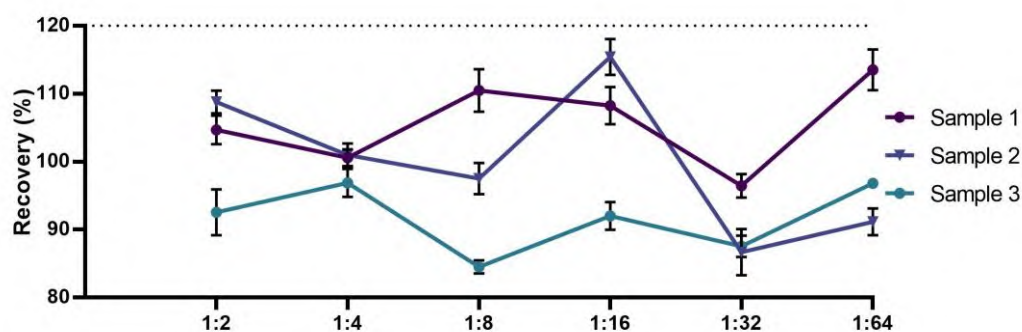
3. Intra-Assay Precision

Ten replicates of each of 5 samples containing different IFN-γ concentrations were tested in one assay. Acceptable criteria: CV < 10%.

Sample Concentration (pg/mL)	Mean (pg/mL)	SD	Numbers	CV
1000	962.03	51.37	10	5.34%
7500	696.46	24.72	10	3.55%
500	458.01	17.59	10	3.84%
15	17.55	1.38	10	7.89%
5	5.41	0.43	10	8.03%

4. Inter-Assay Precision

Five samples containing different concentrations of IFN-γ were tested in independent assays. Acceptable criteria: CV<15%.

| Sample Concentration (pg/mL)	Mean (pg/mL)	SD	Numbers	CV
1000	966.08	45.23	9	4.68%
7500	800.72	41.47	9	5.18%
500	508.73	23.15	9	4.55%
15	14.66	1.11	9	7.60%
5	4.56	0.37	9	8.06%

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5. Recovery

Recombinant IFN-γ was spiked into 5 human serum samples, and then analyzed. The average recovery of IFN-γ for serum samples is 97.76%.

Sample ID	Conc Measured (pg/mL)	Conc Added (pg/mL)	Conc Recovered (pg/mL)	Recovery
1	362.14	400	357.04	89.26%
	269.35	300	264.25	106.76%
	191.15	200	186.05	90.35%
	5.37			
2	371.27	400	366.84	91.71%
	311.4	300	306.97	102.32%
	197.16	200	192.73	96.37%
	4.66			
3	422.46	400	420.52	105.13%
	261.16	300	259.22	86.41%
	228.46	200	226.52	113.26%
	2.04			
4	363.65	400	361.35	90.34%
	301.94	300	299.64	99.88%
	178.21	200	175.91	87.96%
	2.42			
5	422.06	400	417.44	104.36%
	292.74	300	288.12	96.04%
	199.28	200	194.66	97.33%
	4.86			

6. Hook Effect

Not be affected by the concentration of IFN-γ up to 250 ng/mL.
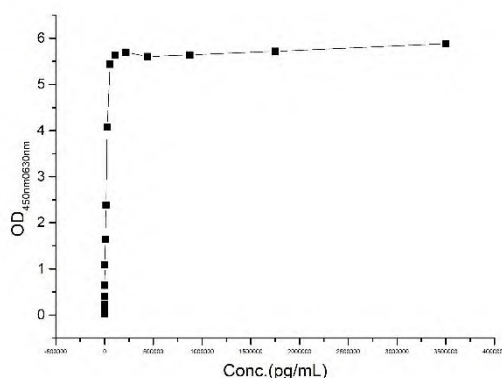
7. Interference Effect

To evaluate the hemolysis matrix effect and high-dose triglyceride matrix effect of assay, serum samples spiked with high concentrations of hemoglobin (2%), Bilirubin (20 mg/dL) or triglyceride (3 mg/mL) were tested. Results shown that all spiked analytes had recoveries between 91% and 103%, no hemolysis matrix effect and high-dose triglyceride matrix effect was observed in assay.

Spiked Material	ID	Conc-1 (pg/mL)	Conc-2 (pg/mL)	Mean (pg/mL)	Recovery
2% Hemoglobin (v/v)	Sample 1	96.44	90.82	93.63	91%
	Spiked Sample 1	82.69	88.51	85.6	
	Sample 2	2.39	2.17	2.28	103%
	Spiked Sample 2	2.38	2.32	2.35	

Spiked material	ID	Conc-1 (pg/mL)	Conc-2 (pg/mL)	Mean (pg/mL)	Recovery
Triglyceride (3 mg/mL)	Sample 1	183.31	179.02	105.95	103%
	Spiked Sample 1	207.12	199.36	108.73	
	Sample 2	143.6	134.5	3.05	101%
	Spiked Sample 2	135.01	127.06	3.07	

Spiked material	ID	Conc-1 (pg/mL)	Conc-2 (pg/mL)	Mean (pg/mL)	Recovery
Bilirubin (20 mg/dL)	Sample 1	183.31	179.02	103.4	98%
	Spiked Sample 1	207.12	199.36	101.56	
	Sample 2	143.6	134.5	2.29	100%
	Spiked Sample 2	135.01	127.06	2.28	

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8. Sample Values

208 healthy serum samples were evaluated for the concentrations of human IFN-γ in assay.
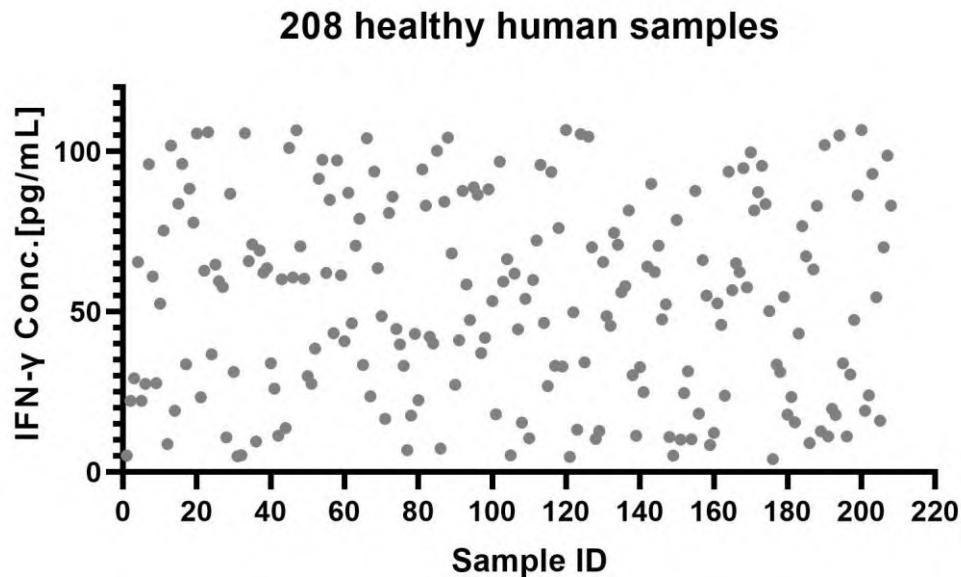
9. Specificity

No cross-reactivity was observed when this kit was used to analyze the following recombinant cytokines at up to 1 μg/mL.

Human	IL-1β, IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-12 p70, IL-10, MCP-1, GM-CSF, TNF-α

10. CALIBRATION

This immunoassay is calibrated against highly purified recombinant human IFN-γ produced at ACROBiosystems. The NIBSC/WHO International Standard for IFN-γ **(82/587)**, which was intended as a potency standard, was evaluated in this kit.

TROUBLESHOOTING GUIDE

Problem	Cause	Solution
Poor calibration curve	* Inaccurate pipetting	* Check pipettes
Large CV	* Inaccurate pipetting * Air bubbles in wells	* Check pipettes * Remove bubbles in wells
High background	* Plate is insufficiently washed * Contaminated wash buffer	* Review the manual for proper wash. * Make fresh wash buffer
Very low readings across the plate	* Incorrect wavelengths * Insufficient development time	* Check filters/reader * Increase development time
Samples are reading too high, but calibration curve looks fine	* Samples contain cytokine levels above assay range	* Dilute samples and run again
Drift	* Interrupted assay set-up * Reagents not at room temperature	* Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of the assay * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts